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RESEARCH ARTICLE

Studies on the Primary Productivity of the Pykara Dam, Nilgiris, TamilNadu, India.

Ramesh. J and M.Lekeshmanaswamy*

PG and Research Department of Zoology, Kongunadu Arts and Science College (Autonomous), Coimbatore-641029, Tamil Nadu, India.

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***Address for correspondence**
Dr. M. Lekeshmanaswamy,
Associate Professor,
PG and Research Department of Zoology,
Kongunadu Arts and Science College (Autonomous),
Coimbatore -641029, Tamilnadu India.
E.Mail:ml_swamy64@yahoo.co.in, mobile-07708570555.

ABSTRACT

Pykara is the name of a village and river located Ooty in the Indian State of Tamil Nadu.The river receives domestic and agricultural wastes and is polluted. Primary productivity is an important phenomenon for aquatic animal. Since the entire heterotrophic community is depended on this [1].As recorded that river become productive when they are enriched with nutrients. The recorded Grossprimary productivity values in the samples of 3 stations ranged from 2.24 to 4.5 mgC/l/hr.The respiratory rate values have been recorded 1.08, 1.03 and 1.03mgC/l/hr.The values of Net productivity has been found to be varied from 0.16 to 1.93 mgC/l/hr in station I, 0.83 to 1.06mgC/l/hr, in station II and 0.50 to 1.07 mgC/l/hr, in station III respectively. The highest productivity observed during summer was probably due to increase in temperature, low turbidity and high transparency.Similar observations has been made by [2] in tropical creek at Gopalpur in Orissa and in the river Cauvery [3]. The observation of low productivity during monsoon could be attributed to a low transparency as reported [4][5][6].Regarding biota of the dam, very limited planktons were identified.

Key words: Primary Productivity, Respiratory activity, Net productivity of Plankton, heterotrophic community.

INTRODUCTION

Pykara is the largest river in the Nilgiri district, TamilNadu State, India. It is considered as a very sacred by the Todas. It is situated about 21 km from Ooty. It is well protected fenced sholas, todas settlement undisturbed grass meadows and also a good wild life habitat. The pykara river orginates at mukurti peak. It passes through hilly track, generally keeping to north and turns to west after reaching the plateaus edge. It gets down majestically in a series of cascades and last two falls of about 55 meters and 61 meters are known as pykara falls. The pykara dam has been constructed and the pykara power plant is the highest head plant so far installed in the whole of Asia. A wonderful boat house is being maintained at Pykara dam by the TTDC.In the Pykara dam, electricity is generated with the help of power plant. After generating electricity the water from the dam flows in to Pykara river. Then the river receives domestic and agricultural wastes and is polluted. Sandynalla is a tributary of Pykara river. BPI, Sandinalla extracted products such as gelatin, ossein and allied chemical products as wastes. It was stepped in the year 1971 in sandynalla near Udagamandalam under private sector. The effluents from this factory have been discharged into the river.

Primary productivity it is associated with the producers which are autotrophic, most of which are photosynthetic, and to a much lesser extent the chemosynthetic microorganisms.These are the green plants, higer macrophytes as well as lower forms, the phytoplankton and some photosynthetic bacteria. Primary productivity is defined as "the rate at which radiant energy is stored by photosynthetic and chemosynthetic activity of producers." Primary productivity is further distinguished as; Gross primary productivity. It is total rate of photosynthesis including the organic matter used in respiration during the measurement period. This is also sometimes referred to as total (gross) photosynthesis or total assimilation. It depends on the chlorophyll content as, Chl/g dry weight/unit area, or photosynthetic number amount of CO2 fixed Gchl/hour. In Net primary productivity. It is the rate of storage of organic matter in plant tissues in excess of the respiratory utilization by plants during the measurement period. This is thus the rate of increase of biomass and is also known as apparent photosynthesis or net assimilation. Thus net primary productivity refers to balance between gross photosynthesis and respiration and other plant losses as death.

MATERIALS AND METHODS

Primary production of the river was measured by light and dark bottle method. The winkler's method of determining dissolved oxygen is normally used in the light and dark bottle technique for productivity rates. The light and dark bottles were suspended in the water by means of strings. At the time of the exposure, the oxygen was fixed in the initial bottle. Then the samples (light bottle and dark bottle) were allowed to remain in the water for an hour for photosynthesis and respiration to take place. After the incubation period the light bottle and dark bottle were taken out from the water and the oxygen was fixed. By following the Winkler's method dissolved oxygen was estimated in the first initial light bottle and dark bottle. The primary productivity can be expressed in terms of oxygen evolved or organic carbon synthesized per unit of water in time't'.

RESULTS AND DISCUSSION

Primary productivity of aquatic ecosystem gives the quantitative details regarding energy fixation and its availability to support bio activity of the total system. The primary production involved in chemo autotrophic process forms the base of energy flow in the ecosystems. It means the food chains and food web of that ecosystem. The daily and seasonal carbon flow in the system forms the base of the annual food pyramid and can be used to estimate the production at higher tropic levels. Increasing anthropogenic activities like washing of clothes, bathing of animals, discharge of domestic wastes. Industrial effluents etc. in and around the aquatic system and their catchment areas have largely contributed to deterioration of water quality leading to their accelerated eutrophication. Productivity of the lake depends on the presence of plankton biomass, enrichment of nutrients and dissolved matter in the water bodies affect diversity of plankton and also physico–chemical properties of water. Diversity in the distribution, abundance and variation in the biotic factors provide information of energy turn over in aquatic ecosystem. A lot of

workers has studied the influence of nutrients and physico-chemical -factors on algal density [7][8][9][10][11][12]. Higher concentration of total suspended solids and total dissolved solids have adversely affected the macro benthic fauna of lakes and this condition may affect the productivity nature of the dam. The fluctuations of productivity values in Pykara Dam in different months were directly or indirectly influenced by environmental factors.

The results of primary productivity have been presented in tables 1, 2 and 3. The values of Gross productivity in station I was ranged from 1.53 to 3,37 mg C/l/hr and respiratory activity from 1,03 to 2.24mg C/l/hr and Net productivity from 0.50 to 1.0-8mgC/l/hr .In station II The values of Gross productivity was ranged from 1.55to 3.35mgC/l/hr, respiratory activity ranged from 1.03to 2.25 mgC/l/hr and net primary productivity was from 0.51 to 1.06 mgC/l/hr In station III. The values of Gross primary productivity was ranged from 2,24to 2,57 mg C/l/hr, respiratory rate from 1.08 to 3.75mgC/l/hr. and Net productivity from 0.16to1.97mg C/l/hr.

CONCLUSION

The higest productivity observed during summer is probably due to increase in temperature, low turbidity and high transparency.Similar observation has been made Hota *et al.,* (1983)[2].The observation of low productivity in the present study during monsoon control attributed to low transparency.

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Table 1.Data showing the Primary productivity of Pykara river during the period of one year (September 2012 to August 2013) at Station I.

Month 2012-2013	Gross productivity (mg C/ l / h)	Respiratory activity (mg C/ l / h)	Net productivity (mg C/ l / h)
Sep.	2.84	1.80	1.04
Oct.	2.55	1.51	1.04
Nov.	2.59	1.76	0.83
Dec.	1.55	1.04	0.51
Jan.	1.81	1.03	0.78
Feb.	2.08	1.25	0.83
Mar.	2.59	1.75	0.84
Apr.	3.07	2.01	1.06
May.	3.35	2.25	1.10
Jun.	2.57	1.54	1.03
July.	2.05	1.20	0.85
Aug.	2.35	1.45	0.90

Table 2.Data showing the Primary productivity of Pykara dam during the period of one year (September 2012 to August 2013) at Station II.

Month 2012-2013	Gross productivity (mg C/ l / h)	Respiratory activity (mg C/ l / h)	Net productivity (mg C/ l / h)
Sep.	2.89	1.82	1.07
Oct.	2.58	1.50	1.08
Nov.	2.62	1.79	0.83
Dec.	1.53	1.03	0.50
Jan.	1.85	1.05	0.80
Feb.	2.09	1.23	0.86
Mar.	2.60	1.74	0.86
Apr.	3.07	2.02	1.05
May.	3.37	2.24	1.13
Jun.	2.59	1.53	1.06
July.	2.07	1.21	0.86
Aug.	2.39	1.46	0.93

Table 3. Data showing the Primary productivity of Pykara dam during the period of one year (September 2012 to August 2013) at Station III.

Month 2012-2013	Gross productivity (mg C/ l / h)	Respiratory activity (mg C/ l / h)	Net productivity (mg C/ l / h)
Sep	3.52	2.71	0.81
Oct.	3.48	2.65	1.83
Nov.	3.72	2.85	1.97
Dec.	2.24	1.08	1.16
Jan.	2.91	1.21	1.70
Feb.	3.04	1.55	1.59
Mar.	3.28	1.65	1.63
Apr.	4.31	3.75	0.56
May.	4.57	3.45	1.12
Jun.	3.61	2.81	0.80
July.	3.54	2.55	0.99
Aug.	3.12	2.06	0.16

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RESEARCH ARTICLE

Biodegradation of Diesel by *Aeromonas hydrophila.*

Lakshmi Priya .J

PG and Research Department of Microbiology, K.S.Rangasamy College of Arts and Science, Tiruchengode -637 215, TamilNadu,India.

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***Address for correspondence**
Lakshmi Priya .J
K.S.Rangasamy College of Arts and Science
PG and Research department of Microbiology,
K.S.Rangasamy College of Arts and Science,
Tiruchengode - 637 215, TamilNadu,India.
Email: jlakshmipriyamicro@gmail.com

ABSTRACT

Hydrocarbon degrading microorganisms play a major role in the environment.The purpose of the present study was to enumerate of Aeromanas sp. from oil contaminated soil and to study degrading capacity, emulcification activity and production of biosurfactant. The hydrocarbon substrate specificity test shown that diesel is also one of the best substrate for growth and emulsification of biosurfactant by Aeromonas. Among 6 strains of Aeromonas, frist strain (A1) shows maximum degradation rate at end of 168 hrs upto19.37% followed by other strains, about 75% of diesel was degraded by Aromonas over a period of 7days. Emulsification upto 75% by A1 followed by A2(63.75%) ,A3(57.5%), Biosurfactant production by A1 strain 0.064g/1 followed by other strains it represents a new type of biosurfactant with strong emulsifying ability.

Key words: Aeromonas, Biosurfactant, Diesel, emulsification, Hydrocarbon.

INTRODUCTION

Biodegradation is the partial or complete conversion of the compound of interest to its elements. The role of organisms, both micro-and macro-organisms, in biodegradation is complex. It is a function of the organism's presence in an environment, their ecology, their metabolism (enzyme complement and efficiencies), growth rate and kinetics (of both growth and metabolism). It has been known for several decades that microorganisms possess both aerobic and anaerobic degradation. Microorganisms are actively involved in the degradation of several naturally occuring and toxic substances such as petroleum hydrocarbons, pesticides etc., Aerobic biodegradation is the breakdown of organic contaminants by microorganisms when oxygen is present. Aerobic bacteria use oxygen as an electron acceptor, and break down organic chemicals into smaller or organic compounds, often producing carbon dioxide and water as the final product. Aerobic biodegradation is also known as aerobic respiration. Aerobic biodegradation is an important component of the natural attenuation of contaminants at many hazardous waste sites. Anaerobic biodegradation is the breakdown of organic contaminants by microorganisms when oxygen is not present. Some anaerobic bacteria use nitrate, sulfate, iron, manganese, and carbon dioxide as their electron acceptors. And break down organic chemicals into smaller compounds, often producing carbon dioxide and methane as the final products. Anaerobic biodegradation is an important component of the natural attenuation of contaminants at many hazardous waste sites.

The most striking feature of a survey of the microorganisms involved in biodegradation processes is their large numbers, ubiquitous presence and varied capabilities. Rather that list the organisms involved (including the bacteria, fungi, actionmycetes, protozoa, etc.) a very brief treatment of the reasons for this versatility is given. Microorganisms as a group show a very wide tolerance range for environmental factors – very low to very high pH levels, 0^0C to 80^0C temperature, of the reasons for this versatility is given.

Biodegradation of organic compounds is the partial simplification or complete destruction of their molecular structure by physiological reactions catalyzed by microorganisms (Alexander, 1981; Atlas and Bartha, 1992; Young, 1997). Biodegradation is routinely measured by applying chemical and physiological assays to laboratory incubations of flasks containing pure cultures of microorganisms, mixed cultures, or environmental samples (e.g. soil, water, sediment, or industrial sludges). Oxygen is one of the most important requirements for microbial degradation of hydrocarbons. However, its availability is rarely a rate-limiting factor in the biodegradation of marine oil spills. Microorganisms employ oxygen-incorporating enzymes to initiate attack on hydrocarbons. Anaerobic degradation of certain hydrocarbons (i.e., degradation in the absence of oxygen) also occurs, but usually at negligible rates. Such degradation follows different chemical paths, and its ecological significance is generally considered minor. Studies of sediments impacted by the Amoco Cadiz spill found that, at best, anaerobic biodegradation is several orders of magnitude slower than aerobic biodegradation (Ward et al., 1980).

Hydrocarbons and its impact on the environment

Petroleum hydrocarbons existed long before humans developed the technological ability to retrieve it from the earth and use it as a source of energy. Natural seeps with in the ocean floor have been releasing the hydrocarbons for thousands of years creating ecosystems with adaptive microorganisms that utilize petroleum effectively. However, the ecological balance in environments those are not adjusted to assimilating large amount of spills or released from large quantity transportation and extraction practices.

Since the mid-1980's, hydrocarbon contamination has become a critical environmental issue in the world due its adverse environmental and health effects. Hence, increasing attention is being given to the study and development of techniques for cleaning up this contamination. To understand the potential environmental impact that can occur during the oil spill, the molecular components of petroleum hydrocarbons must be considered. The natural composition of petroleum diesel is very complex.

Petroleum hydrocarbon continues to be used as the principle source of energy and hence an important global environmental pollutant. Apart from accidental contamination of ecosystem, the vast amounts of oil sludge generated in refineries from water oil separation system and accumulation of waste oily materials in crude oil storage tank bottoms pose great problems because of the expensive disposal methods. (Ferrari et al., 1996; Vasudevan and Rajaram, 2001). Despite decades of research, successful bioremediation of petroleum hydrocarbon contaminated soil remains challenge.Concentration of inorganic nutrients often limits the biodegradation of petroleum hydrocarbons in marine environment. Atlas and Bartha (1972) found that microbial degradation and mineralization were not increased by nitrate or phosphate alone but were increased dramatically when nitrate and phosphate were added together.

Distribution of hydrocarbon degrading microorganisms

It has been known for several years that certain microorganisms are able to degrade petroleum hydrocarbons and use them as sole source of carbon and energy for growth. The ability to degrade and/or utilize hydrocarbons substrates is exhibited by wide variety of bacterial genera, 25 genera of hydrocarbon degrading bacteria (Floodgate, 1984) have been isolated from the marine environment. Similarly 22 genera of hydrocarbon degrading bacteria have been reported (Bossert and Bartha, 1984) based on the number of published.

Microbial remediation of toxic hydrocarbon contaminated sites is carried out by a diverse group of microorganisms. Study of this diversity at the genetic level is necessary to understand the phylogenetic perspective, the mechanism of degradation, and develop novel strategies of treatment. Analysis of microorganisms having high specificity for recalcitrant compound. Documentation of this microbial diversity from oily sludge/crude oil contaminated sites is essential because they create a major environmental concern and these microbes can be used for cleaning up the same.

Microbial degradation of oil has been shown to occur by attack on aliphatic or light aromatic fractions of the oil, with high molecular weight aromatics, resins, and asphaltenes considered to be recalcitrant or exhibiting only very low rates of biodegradation (Joseph et al., 1990) Broderick and Coony (1982) reported that 96% of hydrocarbon utilizing bacteria isolated from freshwater lakes were able to emulsify kerosene, and it has been observed that mixed cultures of marine and soil bacteria which effectively degrade crude oil also exhibit strong emulsifying activity. Both aerobic and anaerobic biodegradation have been shown to reduce the concentration of several components of petroleum hydrocarbons.Mixed cultures often involve significant degradative capabilities because the single strains can complement to one another due to their physiological properties. Therefore, some members of the culture might be able to provide important degradative enzymes whereas other supply surfactants or growth factors.

During oil biodegradation, oil fluid properties change because different classes of compounds in petroleum have different susceptibilities to biodegradation. The early stages of oil biodegradation are characterized by loss of n-paraffin followed by loss of acyclic isoprenoids. Compared with those compounds groups, other compound classes are more resistant to biodegradation. However, even those more resistant compound classes are eventually destroyed as biodegradation proceeds. Hydrocarbons are natural products as well as pollutants; it is not surprising that hydrocarbon oxidizing bacteria, fungi and algae are distributed widely in nature. A typical soil, beach sand or ocean sediment contains 10^4 to 10^6 hydrocarbon degrading microorganisms per gram of soil (Rosenberg, 1991).

Characteristics of diesel

One can obtain diesel from petroleum, which is sometimes called petrodiesel when there is a need to distinguish it from diesel obtained from other sources. As a hydrocarbon mixture, it is obtained in the fractional distillation of crude oil between 250^0C and 350^0C at atmospheric pressure. Diesel is generally simpler to refine than gasoline and often costs less (through price fluctuations often mean that the inverse is true).

Reducing the level of sulfur in diesel is better for the environment. It allows the use of catalytic diesel particulate filters to control diesel particulate emissions, as well as more advanced technologies, such as Nox absorbers (still under development), to reduce emissions of nitrogen oxides (Nox). However, lowering sulfur also reduces the lubricity of the fuel, meaning that additives must be put into the fuel to help lubricate engines. Diesel contains approximately 18% more energy per unit of volume than gasoline, which along with the greater efficiency of diesel engines contributes of fuel economy (distance traveled per volume of fuel consumed). In the maritime field various grades of diesel fuel are used.

Role of Aeromonas in the degradation of hydrocarbons

Aeromonas is a non motile, gram negative rod shaped bacteria, chemoorganotrophic facultative anaerobes demonstrating both respiration and fermentative metabolism. Although it is a pathogen to human as well as fish, 29.6% of total population of Aeromonas was reported in soil followed by Pseudomonas and Bacillus.

Like Pseudomonas and Bacillus sp. Aeromonas also play a vital role in hydrocarbon degradation and production of biosurfactant. The peak growth and biosurfactant production was the 8th day (Ilori et al., 2005). Hydrocarbon degrading microorganisms plays a major role in the environment. Crude oil degrading bacterial strains were isolated from refinery oil contaminated soil which includes the Aeromonas sp. the rate of degradation at the 8th day 53.55% (Vivekanandhan et al., 1999).Several studies on production of biosurfactants by Aeromonas sp. were also reported (Desai et al., 1987; Lang et al., 1987; Rosenberg, 1986; Wilkinson et al., 1985).

Biosurfactants

Classification of biosurfactants

Biosurfactants can be classified in several broad groups: Glycolipids, lipopeptides, lipopolysaccharides, phospholipids, fattyacids, and neutral lipids. The classification of biosurfactants is based on their chemical nature. Low molecular weight substances (e.g., rhamnolipid phospholipids, peptides), polymeric materials (e.g., proteins, polysaccharides) or particulate compounds (e.g., extracellular vesicles or microbial cells) (Rosenberg, 1986)

Production of biosurfactants by microorganisms

Biosurfactants, which are natural emulsifiers of hydrocarbons, are produced by some bacteria, fungi and yeast. Biosurfactant is defined as a surface active molecule containing both hydrophobic and hydrophilic components which is produced by microorganisms.

Biosurfactants can improve the bioavailability of hydrocarbons to the microbial cells by increasing the area at the aqueous hydrocarbon interface. This increases the rate of hydrocarbon dissolution and thereby utilization by microorganism (Gerson, 1993). Surface active biosurfactants are employed for enhanced oil recovery (Hart, 1989) and as flocculating agents, as detergents and adhesives (Zajie and Saffens, 1984).

Application of bio-surfactants

Research in the area of biosurfactants has expanded quite a lot in recent years due to its potential use in different areas, such as the food industry; agricultural, pharmaceutical, oil industry, neurochemistry and the paper and pulp industry. The development of this line of research is of paramount importance, mainly in view of the present concern with protection of the environment. Therefore, the most significant advantage of microbial surfactants over chemical surfactants is its ecological acceptance because of its biodegradability and nontoxic to natural environments. The emphasis to date has been on enhanced oil recovery, cleaning oil spills, oil emulsification and in breaking industrially

derived oil-in-water and water-in-oil emulsions. Dispersion and solubilization of organic compounds having low water solubility is an important step in bioremediation. Biosurfactants offer potential for dealing with this problem by increasing the dispersion and solubilization of organic compounds having limited water solubility.

Biosurfactants have been used for gene transfection, as ligands for binding immunoglobulins, as adjuvants or antigens and also as inhibitor for fibrin clot formation and activators of fibrin clot lysis. Biosurfactants have the potential to be used as a preventive strategy to delay the on set of pathogenic biofilm growth on catheters and other medical insertional materials, thus lowering the large number of hospital infections without the use of synthetic drugs and chemicals. In spite of the immense potential of the biosurfactants, their use still remains limited, because of their comparatively high production cost, as well as scant information on their toxicity towards human systems. However, it is only a matter of time before the full potential of biosurfactants is fully exploited and used in medical science (Karnath et al., 1999).

Importance of the Present Investigation

Petroleum hydrocarbon continues to be used as the principle source of energy. Wide scale production, transport use and disposal and petroleum globally have made it a major containment in both prevalence and quantity in the environment. Biosurfactant are a group of surface-active molecules produced mainly by hydrocarbon degrading microorganism it can degrade or transform the components of petroleum products. They are non-toxic, non–hazardous, Biodegradable and environmentally friendly components. Hence, reclamation of petroleum hydrocarbon polluted sites can be carried out by bioremediation, which is can enhance natural process of biodegradation using biosurfactant producing and oil degrading bacterial cultures. Bioremediation technologies generally aim at providing favourable conditions of certain, temperature and nutrients to enhance biological hydrocarbon break down.

Most work on biosurfactant production by microorganisms has been concentrated on the determination of the distinct polar moieties used for categorization into classes such as glycolipids and lipoproteins. While biosurfactant production by organisms such as Pseudomonas sp. Achinetobacter sp. Bacillus sp. Rhodococcus sp. and Arthobacter sp. have been well studied, only a few reports exist on the ability of Aeromonas sp. to the best of our knowledge, to produce biosurfactant when grown on hydrocarbon. Among the soil microorganisms which include Bacillus and Pseudomonas, the presence of Aeromonas ranges about 29%. Apart from its pathoginicity in humans and fish, its also play a vital role in hydrocarbon degradations.

The present study is designed to carry out the following objectives.

- To enumerate of Aeromonas sp. from oil contaminated soil.
- To study the degradation capacity of *Aeromonas sp*.
- To study the emulsification activity.
- To estimate the biosurfactant production.

MATERIALS AND METHODS

Collection of soil samples

The oil contaminated soil samples were collected from petrol station at Tiruchengode in a sterile polythene bag and transported to laboratory within one hour. The samples were processed by enrichment method to enumerate oil degrading Aeromonas sp.

Enrichment method

One hundred millilitres of Mineral Salts Medium (MSM) was distributed in flasks and 1 g of soil was added into the medium followed by the addition of 0.5% of diesel as carbon source. The flask was kept in the shaker (110 rpm) at room temperature for uniform distribution of diesel. After 24 – 48 hrs 1 ml of the liquid culture was transferred to fresh MSM for second enrichment and 0.5% diesel was again added as carbon source and the flasks were kept in a shaker at room temperature for 24 – 48 hrs. After incubation, samples were serially diluted and plated over nutrient agar medium and incubated at room temperature for 24 – 48 hrs. Morphologically different colonies were isolated and considered as hydrocarbon utilizing organisms.Among which, Aeromonas colonies were selected and transferred to mineral salt medium (MSM), supplemented with 0.5% of diesel. After 24 – 48 hrs of incubation a loopful of liquid culture was streaked on agar plates and sub-cultured on fresh agar plates, in order to get the pure culture of Aeromonas. The isolates were maintained on nutrient agar slants at 4^0C and sub cultured every 2 weeks.

Identification and characterization of Aeromonas sp.

The morphological and biochemical tests as per Bergey's Manual of Systematic Bacteriology (1994).

Growth substrate range determination (Ilori et al., 2005)

The ability of the isolate to utilize diesel as sole carbon source pf carbon and energy was determined. The carbon source (0.5%, vv^{-1}) was added to MSM (100ml) contained in 250ml Erlenmeyer flasks. A non-inoculated control flask was prepared for comparison purpose. The media, after sterilization, were inoculated with test organism. Incubation was with shaking (120 rpm) at room temperature (30 ± 2^0 C) for 7days.

Composition of media

The mineral salts medium (MSM) (Kastner et al., 1994) containing the following composition was used.

Disodium hydrogen phosphate	-	2.13g
Dipotassium hydrogen phosphate	-	1.3g
Ammonium Chloride	-	0.5g
Magnesium Sulphate	-	0.2g
Distilled water	-	1L
pH	-	7

Composition of trace elements (Bauchop and Elsden, 1969)

Boron	-	0.025%
Copper Sulphate	-	0.05%
Manganese Sulphate	-	0.05%
Molybdenum Chloride	-	0.006%
Zinc Sulphate	-	0.07%
Distilled Water	-	100ml3

Estimation of biomass (Rahman et al., 2002)

3 ml of the samples were withdrawn from the culture flasks two days interval and centrifuged at 4000rpm for 20minutes.The pellet containing cells was dried in an oven at 110^0 C and the biomass was calculated.

Emulsification (Iqbal et al., 1995)

The ability to emulsify liquid hydrocarbons, diesel was determined. The 2ml broth was added into each test tube (diesel 2ml). The content of the tubes were vortexed at high speed for 2min and left undisturbed for 24h. The emulsion index was determined as the height of the emulsion layer divided by the total height and multiplied by 100.

Surface tension measurement (Lang and Wanger, 1987)

Surface tension is defined as the force acting perpendicular to a line of unit length drawn on an imaginary plane of film of a liquid or in other terms the force capable of, to bind the drop, of liquid together in itself to give the maximum possible size if it could hang by its own weight under gravity. This definition also leads to its measurement by simple technique called drop weight method. By this method, a vertical fine capillary nozzle having round tapered mouth is required. The liquid is passed slowly to make a fine drop which hangs by its own weight and then falls down by gravity. The weight of a single drop from cell free supernatant was measured by taming average of several statistical weight data of 200 drops for each sample.

The following emperical formula was applied to calculate the surface tension in mN/m.

$$T = \frac{mxg}{3.8xr}$$

Where,

m = mass of a single drop of liquid (kg)
r = outer round tapered radius of the nozzle (m)

Biosurfactant estimation (Zhang et al., 1992)

Cell-free culture broth (1ml) was added to 4.5ml of dilute sulfuric acid (6:1v/v) and mixed thoroughly. The mixture was heated at 100^0C for 10min and cooled to room temperature. To the mixture was added 0.1ml of freshly prepared 3% solution of thioglycolic acid, and the mixture was incubated in darkness for 3h. Absorbance was measured at 400 and 430nm spectrophotometrically. Rhamnolipid concentration was calculated using the formula

$$RL = [54.18 (A_{400} – A_{430}) – 1.49]F$$

Where A_{400} and A_{430} are absorbances at 400 and 430nm, respectively, and F is the dilution factor. A standard curve prepared using different concentrations of L-rhamnose (Sigma) was used to determine the rhamnolipid concentration.

RESULTS AND DISCUSSION

Enumeration of bacterial population

The total heterotrophic bacterial population (THB) from oil contaminated soil samples was enumerated. After the enrichment process, 1ml of enriched culture was serially diluted and plated using pour plate technique. The THB population was ranged between $1.6x10^5$ and $3.4x10^6$cfu/g whereas in uncontaminated soil, the THB population may vary between 10^7~10^9cfu/g. This variation is due to the presence of hydrocarbon in the contaminated soil. As per the studies of Rahman et al (2003) the size of the microbial population decrease is based on the chemical composition of contaminating oil and species of microorganisms present in the microbial community of particular ecosystem. Plate

counts confirm the presence of a significant number of hydrocarbon-oxidizing organisms in soil. Morphologically different isolates were observed on the nutrient agar plates.

As it was aimed to work on the degradative potential of Aeromonas species, oxidase and catalase positive isolates were subjected to biochemical analysis. Gram-negative rod, motile, urease negative, it utilized lysine, citrate and produced gas from glucose. Maltose and lactose were not utilized. The organism reduced nitrate, utilized ornithine, but not sorbitol. It was therefore putatively classified as the species of Aeromonas, taken for the present investigation. In unpolluted ecosystem, hydrocarbon utilizers generally constitute less than 0.1% of the microbial community and in oil polluted ecosystems they can constitute upto 100% of the viable microorganisms. The microbial populations quantitatively reflect the degree or extent of exposure of that ecosystem to hydrocarbon contamination (Atlas 1981, Al- Gounaim et al., 1995).

The rapidly expanding literature on the oxidation and assimilation of hydrocarbon substrates by soil microorganisms attests to the widespread occurrence and ease of isolating these organisms from nature (McClay et. al., 2000 and Van Dyke, 1991). Population levels of hydrocarbon utilizers and their population within the microbial community appear to be a sensitive index of environmental exposure to hydrocarbons.Microorganisms are known to attack specific compounds present in crude oil that is a complex mixture of saturates, aromatics and polar compounds (Bharathi and Vasudevan, 2001). An effective degradation of crude oil would require simultaneous action of several metabolically versatile microorganisms with favorable environmental conditions such as pH, temperature and availability of nutrients (Venkateswaran and Harayama, 1995). An oil spill in the environment leads to an adaptive process and if metabolically active hydrocarbon utilizing microorganisms could respond would be reduced considerably. The necessity for seeding with complementary population may not be capable of degrading a wide range of potential substrates in a complex mixture such as crude oil (Chhatre et al., 1996).

Growth substrate range determination

The results of the hydrocarbon substrate specificity test revealed that the organism had good growth on diesel as substrate. When no difference was noticed in the turbidity of the test flask and that of the control, it was taken as no growth (-), when slight increase in turbidity was noticed, it was taken as poor growth (+). Significant increase in turbidity was taken as good growth (++).

In the present study among six strains of Aeromonas, four strains shows increase in turbidity in the presence of diesel as substrate was taken as good growth (++) and two strains shows slight turbidity was taken as poor growth (+). Thus the substrate specificity test conclude that diesel also act as best substrates for growth of hydrocarbon degraders (Table 1).The addition of hydrocarbons to an ecosystem may result in a selective increase in microorganisms capable of utilizing the hydrocarbons and those that are capable of utilizing metabolites produced by the hydrocarbon-utilisers (Venkateswaran and Harayanma, 1995, Ferrari et al., 1996). The enhancement or reduction will depend upon the chemical composition of the contaminating hydrocarbons and on the species of microorganisms present within the microbial community of the particular ecosystem (Atlas, 1995).The added oil enriched for the species that have inherent petroleum hydrocarbon assimilating potential (Bossert and Bartha, 1984), whereas the less adapted species among the total heterotrophic population are gradually eliminated, resulting in qualitative shifts in species composition (Amadi, 1990).

Degradation of diesel

Hydrocarbon degradation by microbial communities depends on the composition of the community and its adaptive response to the presence of petroleum hydrocarbon. The organism used in this study was isolated from oil contaminated soil samples from petrol station at Tiruchengode, the organism therefore might have had prior exposure to hydrocarbons like diesel, thus it shows good growth in diesel.

In the present study the bacterial strain Aeromonas developed from oil polluted sites grown well on diesel .The rate of degradation is proportional to the bacterial population. The degradation of diesel was observed at every 24hrs interval and there was a corresponding increase in the bacterial cell population. The degradation of diesel after 24hrs was 2.57% at the end of 168 hrs strain A1 degraded upto 19.37%, shows maximum degradation followed by the other strains (Table 2).

The rate of microbial degradation of hydrocarbons in soils is affected by several physico-chemicals and biological parameters including the number and species of microorganisms present, the conditions for microbial degradation activity (e.g. presence of nutrient, oxygen, pH and temperature) the quality, quantity and bioavailability of the contaminants; and the soil characteristics such as particle size distribution (Margesin and Schinner, 1997).Isolation of degrading strains was performed with diesel oil, crude oil as sole carbon source, the result showed a reduction of 30% while this value decreased to 0.15% in uncontaminated samples, which includes the strains of Aeromonas sp and Pseudomonas vasicularis (Saagua et al., 2002).

Petrodiesel oil degraders belong to the genera Micrococcus, corynebacterium, Bacillus, Entrobacter, Pseudomonas, Alcaligenes, Flavobacterium, Moraxella, Aeromonas, Acinetobacter and Vibrio. The flora reflects the normal hetrotropic bacteria present in soil and native genera seem to be crude oil utilisers. Several other workers also reported on the above genera as hydrocarbon degrading microorganisms (Atlas 1981, Leahy and Colwell 1990, Banat et al ., 2001).The rate of hydrocarbon degradation ranged from 0 μ gg^{-1} to 0.60 μgg^{-1} for control oil, 0.05 μgg^{-1} to 1.67gasoline and from 0.12 μgg^{-1} diesel oil to 1.31 μgg^{-1} for hydrocarbon contaminated soils respectively (Obire, 2001). The biodegradability of seven different crude oils was found to be highly dependent on their composition and on incubation temperature. At 20^0C lighter oils had greater abiotic losses and were more susceptible to biodegradation than heavier oils (Atals, 1995).

Chhatre et al., (1996) reported about 60% of degradation of crude oil using semicontinuous crude oil fed reactor using a four members consortium. Several other workers (Venkateswaran and Harayama 1995, Lal and Khanna, 1996, Sugiura et al., 1997) showed that a bacterial consortium was able to degrade 28-51% of saturate and 0-18% of aromatics present in crude oil or up to 60% crude oil by mixed consortia. The percentage of biodegradation was significantly higher than that achieved by individual isolates.

By the addition of metabolically active hydrocarbon utilizing microorganisms, the lag period before the indigenous microbial population respond to the addition of a complex mixture such as diesel oil can be reduced considerably (Del'Arco and De Franca, 1999; Bharathi and Vasudevan, 2001). Several other workers (Chhatre et al., 1996; Sugiura et al., 1997; Vasudevan and Rajaram, 2001) have described the ability of mixed bacterial consortia to degrade 28-51% of saturates and 0-18% of aromatics present in petrodiesel or upto 60% petrodiesel.

Biostimulation and bioaugmentation on the degradation of total petroleum hydrocarbon (TPH) in soils contaminated by diesel oil, showed the greatest degradation upto 72.7% by the number of diesel oil degrading microorganisms includes Aeromonas, Bacillus, Acinetobacter (Fatima et al., 2003).A survey of soils from the northwest area of Canada for the presence of oil-utilizing microorganisms indicated that not all soils have an indigenous population capable of utilizing oil degradation rates of the bacterial consortium such as Flavobacterium and Cytophaga (41%), Pseudomonas sp. (34%), Xanthomonas (9%), Aeromonas (17%) respectively (Jobson et al., 1972).

Emulsification

The formation of a water-in-oil mixture. An emulsified mixture of water in oil is commonly called mousse. The presence of mousse indicates that a spill has been on the water for some time.

The liquid aromatic hydrocarbons were particularly not good substrates for emulsification, however diesel was found to be good substrates for emulsification in the present study out of 6 strains of Aeromonos A1, A2, A3 shows 75%, 57.5%, 63.75%, respectively found to be good emulsifier (Table 4 and Figure 3). Rosenberg (1994) suggested that the natural role of emulsions is to enhance the growth of bacteria on petroleum hydrocarbons. The ability of the extracellular emulsifying agent of Arthrobacter sp, Aeromonas sp, Pseudomonas sp. has been reported (Rosenberg et al., 1979). Emulsification is known to enhance hydrocarbon metabolism (Berg et al., 1990; Hommel, 1993). Stability of emulsion in the presence of salt has been reported as one of the properties of the biosurfactant produced by Bacillus licheniformis strain (Mclnerney et al., 1990). Broderick and Cooney (1982) reported that 96% of hydrocarbon utilizing bacteria isolated from lakes were able to emulsify kerosene which effectively activity.

Rosenberg et al., (1991) reported that the ability of the extracellular emulsifying agent of Arthrobacter sp, Aeromonas and Bacillus sp. to emulsify crude oil and fractions of crude oil, is, gas oil was a better substrate induced emulsification than kerosene. In fact, emulsions of gas oil were as stable as crude-oil emulsions. Diesel light petroleum has yielded emulsions and the emulsions derived from kerosene and gasoline were unstable. Pentane and hexane also were not emulsified effectively; however quantitative data for these tow paraffins were not obtained because of extensive evaporation during incubation. A higher emulsifying activity has been reported that the biosurfactant produced by Pseudomonas. The emulsions were stable at temperatures ranging from 0^0C to 100^0C (Rosenberg, 1992).

Most microbial surfactants are substrate specific, solubilizing or emulsifying different hydrocarbons at different rates (Ilori and Amund, 2001). An emulsion is formed when one liquid phase is dispersed as microscopic droplets in another liquid continuous phase (Desai and Banat, 1997). Poor emulsification of some of the hydrocarbons might be due to the inability of the biosurfactant to stabilize the microscopic droplets. Emulsifying biosurfactants that are stable in environments with high pH and salinity would find applications for bioremediation of spills at seas. The biosurfactant may also be useful for bioremediation works in hot and slightly alkaline environments.

Surface tension measurement

Surface tension is a measurement of the cohesive energy present at an interface. The molecules of a liquid attract each other. The interactions of a molecule in the bulk of a liquid are balanced by an equal attractive force in all directions.

In the present study, strains of Aeromonas A1, A2, A3, A4, A5, A6 lowered the surface tension of water to 30, 25, 28, 22, 20, 25, 20mN/m respectively (Table 5). Several strains of anaerobic bacteria produce biosurfactnats (Grula et al., 1983). However, the observed reduction in surface tension (45 to 50 mN/m) was not as large as the observed reduction in surface tension by anaerobic organisms (27 to 50 mN/m) (Cooper, 1986). Lowering of surface tension is an important property of hydrocarbon degrading strains which helps in utilization of the hydrophobic substrates (Rahman, 1993) reported a surface tension value of 29.5mN/m with 1% inoculum at the stationary phase. The glycerol medium with 10% inoculum produced the lowest surface tension value of 29.5mN/m. An effective microbially produced surfactant can lower this value to <30dyn/cm (Lang and Wanger, 1987). The lowest value of surface tension by Bacillus sp. and Aeromonas sp. was reported by (Takeyama and Matsunaga, 2002). In a study by Oberbremer and Muller-Hurtig (1989), a positive correlation was obtained between reduction in surface tension of the fluid phase in a stirred soil bioreactor and the onset of biodegradation of hydrophobic petroleum hydrocarbons. It has also been previously reported that rhamolipid can mediate reduction in surface tension (Banat et al., 2000; Noordman et al ., 2000).

Biosurfactant estimation

Biosurfactants, amphiphilic compounds of microbial origin, have advantages over their chemical counterparts in biodegradability and effectiveness at extreme temperature, pH and in having lower toxicity (Banat et al., 2000). In the

present study out of 6 strains of Aeromonas, A1 shows 0.041g/l of biosurfactant production after 24hrs and at 168 hrs 0.064g/l, A2 shows 0.049g/l at 24hrs and at 168 hrs 0.067g/l, A3 shows 0.043g/l at 24hrs and at 168 hrs 0.064g/l, A4 shows 0.038g/l at 24hrs and at 168 hrs 0.056g/l, A5 shows 0.052g/l at 24hrs and at 168 hrs 0.067g/l, A6 shows 0.034g/l at 24hrs and at 168 hrs 0.049g/l respectively among which the maximum biosurfactant production was show by the A1 at 168hours (Table 6 and Figure 3).

Biosurfactants have been reported to be produced on water soluble compounds such as glucose, sucrose, glycerol, or ethanol (Desai and Banat, 1997). Biosrufactant produced from water-soluble substrates have been reported to be inferior to that obtained with water immiscible substrates (Syldatk et al., 1985; Robert et al., 1989). Such biosurfactants may however be cheaper to produce and useful in food and pharmaceutical industries as it will not required extensive purification. Biosurfactants producing microorganisms may play an important role in the accelerated bioremediation of hydrocarbon contaminated sites (Banat et al., 2000; Rosenberg et al., 1999). Most microbial surfactants are substrate specific, solubilizing or emulsifying different hydrocarbons at different rates (Ilori and Amund, 2001). Moreover, use of biosurfactant producing, hydrocarbon degrading, microorganisms for bioaugmentation to enhance hydrocarbon degradation offer the advantage of a continuous supply of a non-toxic and biodegradable surfactant at a low cost (Moran et al., 2000; Rahman et al., 2000c). Environmental factors such as pH, salinity and temperature also affects biosurfactants activity.

Our results indicate that Aeromonas sp. were efficient in biosurfactant production and hydrocarbon emulsification. These results suggests that the use of carbon source like diesel for the biosurcatant production, could enhance the biosurfactant production and there by these strains may be applied for bioremediation of hydrocarbon-contaminated sites and enhanced oil recovery.

CONCLUSION

The discovery of petroleum brought a lot of relief to the world's energy requirement because of ease of sourcing and conversion. The ease of production, refining and distribution has also brought with it an ever-increasing problem of environmental pollution. One of the ways through which petroleum pollutants can be removed is by solubilization and emulsification. Hydrocarbon oxidizing bacteria, fungi and algae are distributed widely in nature. Fertile soil contains significant number of microorganisms that can utilize hydrocarbon as sole source of carbon and energy. As a hydrocarbon mixture, diesel is obtained in the fractional distillation of crude oil between 250^0C and 350^0C at atmospheric pressure. It is generally simpler to refine than gasoline. The purpose of present study was to enumerate the diesel degrading Aeromonas and its emulsification activity and production of biosurfactant. The organism used in this study was isolated from oil contaminated soil; the hydrocarbon substrate specificity test showed that diesel is also one of the best substrate for growth and emulsification of biosurfactant by Aeromonas. Among six strains of Aeromonas, first strain shows maximum degradation rate and emulsification of biosurfactant. About 75% of diesel was degraded by Aeromonas over a period of 7 days. The biosurfactant produced by the diesel degrading Aeromonas sp. represents a new type of biosurfactant with strong emulsifying ability

Identification of Aeromonas strains with proper identity and identification of other microbes which are inhabited in the same natural environment will rive a clear idea about the microbial diversity in the particular automobile service station environment – where petroleum hydrocarbon and detergent levels would be very high. Biosrufactant productivity studies, identification and characterization of biosurfactants by TLC and HPLC will be highly applicable to understand and interaction between microbes and their environment. Emulsification and hydrocarbon degradation studies in the future will give a clear picture about the degradation of petroleum compounds by Aeromonas sp.

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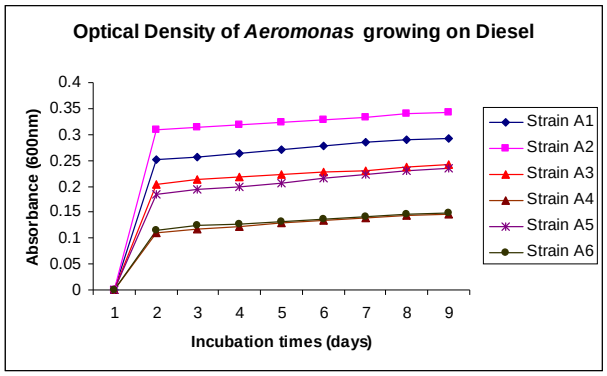

Fig.1: Optical Density of Aeromonas growing on Diesel

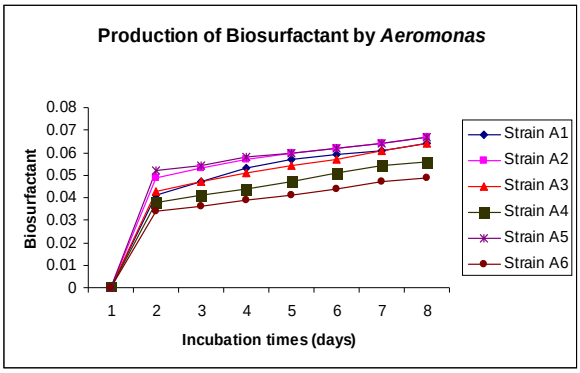

Fig.2: Production of Biosurfactant Aeromonas

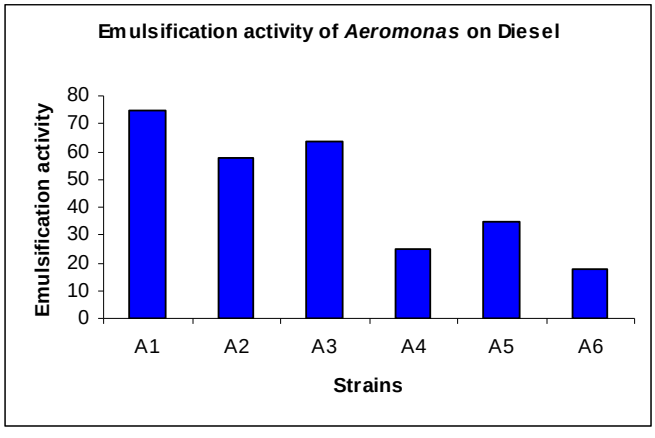

Fig.3: Emulsification activity of Aeromonas on Diesel

Table 1.Growth range determination of Aeromonas

Strains	Turbidity
Control	-
A1	++
A2	++
A3	++
A4	+
A5	++
A6	+

Table 2.Degradation of diesel by Aeromonas

Incubation Periods (Days)	Strains /O.D Value						
	Control	A1	A2	A3	A4	A5	A6
0	0.000	0.251	0.308	0.203	0.111	0.184	0.114
1	0.000	0.257	0.313	0.212	0.117	0.193	0.124
2	0.000	0.264	0.319	0.219	0.121	0.198	0.127
3	0.000	0.271	0.324	0.222	0.129	0.207	0.131
4	0.000	0.279	0.329	0.227	0.133	0.216	0.136
5	0.000	0.284	0.334	0.231	0.139	0.223	0.141
6	0.000	0.289	0.339	0.237	0.143	0.229	0.146
7	0.000	0.293	0.342	0.241	0.147	0.234	0.149

Table 3.Biomass Production by Aeromonas

Incubation Periods (Days)	Strains/ Biomass					
	A1	A2	A3	A4	A5	A6
2	0.05	0.03	0.05	0.02	0.03	0.02
4	0.07	0.05	0.06	0.04	0.04	0.03
6	0.09	0.06	0.08	0.05	0.06	0.04

Table 4. Emulsification activity by Aeromonas

Strains	Hydrocarbon	Percent Emulsified
A1	Diesel	75
A2	Diesel	57.5
A3	Diesel	63.75
A4	Diesel	25
A5	Diesel	35
A6	Diesel	18

Table 5. Surface Tension Measurement

Strains	Surface tension (mN/m)
A1	30
A2	25
A3	28
A4	22
A5	24
A6	21

Table 6. Estimation of biosurfactant by Aeromonas

Incubation Periods (Days)	Strains/OD Values						
	Control	A1	A2	A3	A4	A5	A6
1	0.000	0.041	0.049	0.043	0.038	0.052	0.034
2	0.000	0.047	0.053	0.047	0.041	0.054	0.036
3	0.000	0.053	0.057	0.051	0.044	0.058	0.039
4	0.000	0.057	0.060	0.054	0.047	0.060	0.041
5	0.000	0.059	0.062	0.057	0.051	0.062	0.044
6	0.000	0.061	0.064	0.061	0.054	0.064	0.047
7	0.000	0.064	0.067	0.064	0.056	0.067	0.049

RESEARCH ARTICLE

Comparative Studies on Phytochemical profile of *Spirulina platensis* and *Oscillatoria sp. Invitro*.

Indumathi.A[1]* and R. Balaji [2]

[1]Department of Biotechnology, Bon Secours College for Women, Vilar bypass road, Thanjavur-613 006, Tamil Nadu, India.

[2]Department of Chemistry, Rajah Serfoji Govt. College, Thanjavur-613005, Tamil Nadu, India.

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***Address for correspondence**
Mrs.Indumathi.A,
Assistant Professor and Head,
Department of Biotechnology,
Bon Secours College for Women,
Vilar bypass road, Thanjavur-613 006,
Tamil Nadu, India
E.mail: indubalaji82@gmail.com

ABSTRACT

Cyanobacterial blooms have a wide range of social environmental and economic impacts. They are structurally diverse and widely distributed throughout the world and are later known as blue green algae. They are the rich source for many useful natural products that are used as feed and fertilizer. Blue green algae in general contain a significant amount of Protein and carotenoids, namely beta carotene, lycopene and lutein wich has a great significance in therapeutic field. *Spirulina* for its increased nutritional value has been exploited.*Oscillatoria* can be used as an alternative to feed poultry, animals, fishes etc.

Keywords: Spirulina, Carotenoid, Clorophyll, Blue green algae, phytopigments.

INTRODUCTION

Blue green algae (Cyanobacteria) dating back almost 3.5 billion years, diversified extensively to become one of the most successful and ecologically significant group. They had a great impact on earth's early environment. The cyanobacteria are photosynthetic microorganisms, which inhabit in different and extreme conditions. This indicates a high degree of biological adaptation, which has enabled these organisms to thrive and compete effectively in nature [23]. They form a massive population in water bodies characterized by high levels of carbonates and bicarbonates and high pH (up to 11) [8].They are cosmopolitan inhabiting extreme habitats including deep seavents, hot springs and ice. Algae are rich in proteins, essential amino acids, vitamins and natural Beta -Carotene and minerals such as

zinc, selenium and Magnesium[16].Edible blue green algae like *Spirulina, Aphanizomen* and *Nostoc* are being used as food since early ages.The genus *Oscillatoria* and *Spirulina* comprises many species, which are nonheterocystous, filamentous organisms belonging to the order Oscillatoriales and family Oscillatoriaceae. Cyanobacteria have yielded new types of products, not found in higher plants and traditional drug sources[26].The cyanobacterium *Spirulina* has already been commercially exploited in several countries as health foods and therapeutic preparations because of its valuable constituents particularly proteins and vitamins[3,29].

Recent studies have reported that eating foods containing a high chlorophyll content protects liver cancer [6] increases haemoglobin content[15] and promote healing of skin as well as stomach ulcers[12-14].Beta –Carotene as well as other carotenoids used for years as a poultry, animal and fish feed supplement have been shown to be powerful anti oxidants and helpful in the prevention of cardiovascular dieases[1].Beta -Carotene was also proven to stimulate the immune system [17] and prevent skin[25]oral and breast cancer[11,22]. Phycocyanin, the blue pigment present in all blue green algae serves as a protein storage unit and as an anti oxidant and anti inflammatory agent [24]. It has been proven to have significant anti cancer properties in animal models[10,13]in recent years there has been a tremendous increase in the interest on the value added product of blue green algae. The present study aims at the comparision of the various pigments and protein content of *Oscillatoria Sp.* and *Spirulina platensis invitro*. Realising the need to develop the production of natural products sources, this study could help to assess the ability to scale–up the production of natural products compounds in the field of food industry, animal feed supplement etc., thereby decreasing the cost of raw material/ substrate and hence the overall cost of production.

MATERIALS AND METHODS

Sample collection

The cultures of *Spirulina platensis* and *Oscillatoria Sp.* were obtained from S.S Biotech, *Spirulina* production research and training centre at Sakkimangalam, Madurai. The culture purity was cheked by microscopic observation. A loop full off *Spirulina platensis* was cultured in sterile Zarrouks Media while *Oscillatoria Sp*. was inoculated in sterile CHUs media No.10 and incubated at room temperature under sun light for ten days. After incubation, a drop of each suspension was placed individually in glass slide and observed under microscope.The cultures of *Oscillatoria Sp.* and *Spirulina platensis* grown in synthetic medium were analysed for protein and pigment concentration.

Estimation total protein concentration

The total protein content of both the algae was measured by Lowry's method, [20]. 1ml of the Cyanobacterial suspension was homogenized. Then 4ml of alkaline mixture (prepared by adding 1% CuSo4, 2% Sodium Potassium tartarate and 2% Sodium carbonate in 0.1 M NaOH in the ration of 1:1:100)was added. It was mixed well at room temperature for 15 min. Then 0.5ml of Folin Phenol solution was added and mixed well. It was incubated in dark at room temperature for 10 min. the absorbency was measured at 640nm. The concentration of the protein content of *Oscillatoria Sp*. and *Spirulina platensis* was estimated by comparing the optional density values with the standard graph of Protein.

Estimation of Carotenoid concentration

10ml of homogenous suspension of the algal cutures were collected into respective sterile centrifuge tubes aseptically.The homogenous cyanobacterial suspension was centrifuged at 5000 rpm for 10 min. The pellet was washed twice in distilled water. The pellet was then homogenized with 3ml of 85% Acetone. The contents were centrifuged at 5000 rpm for 5 min. The supernatant was collected and made up to 10 ml with 85% Acetone. The absorbancy was measured at 450nm against the acetone blank for carotenoid estimation using the formula

Beta Carotene concentration =DxVxFx10 / 2500 mg/ml
D= Absorbancy at 450m
V= Volume of the sample
F= Dilution factor
2500= Extinction coefficient.

Estimation of Chlorophyll–a concentration

The gross production can be estimated by the relatively simple procedure of extracting pigments and then measuring the chlorophyll concentration with the spectrophotometer. 10ml of the homogenous suspension of the cyanobacterial culture from the medium was collected in sterile centrifuge tubes aseptically. It was then centrifuged at 5000rpm for 10min. The pellet was washed twice in distilled water. It was suspended in 4ml of 80% Methonol and vortexed thoroughly. The mouth of the test tube was covered with aluminum foil to prevent the solvent evaporation. The tubes were incubated in a water bath at 60◦c for 1 hour (preferably in dark) with occasional shaking. It was cooled and centrifuged at 5000 rpm for 5 min. The supernatant was collected and made up to 10ml with 80% methanol (to compensate the solvent loss during heating). It was read at 660 nm in spectrophotometer against methanol blank. The concentration of Chlorophyll was calculated using the formula

Chlorophyll- a Concentration =A663x 12.63x Vol. of sample/Vol. of Methanol µg/ml
A663= absorbance at 663 nm
12.63= correction error

Estimation of Phycobillin concentration

The phycobillin concentration was estimated by a simple procedure and was measured by spectrophotometer. The homogenous algal suspension of the cyanobacterial culture from the medium was collected into a sterile centrifuge tubes aseptically. The suspension was then centrifuged at 5000 rpm for 10 min. the pellet was washed twice in distilled water. It was then homogenized with 3ml of phosphate buffer solution. The suspension was repeatedly freeze and thawed and centrifuged at 5000 rpm for 5min. the supernatant was collected and the absorbency was measured at 565,615 and 625nm respectively against phosphate buffer blank. The different phycobillin concentration are calculated by using the formula,

C- Phycocyanin (PC): =A615-0.474(A652)/5.34 mg/ml
Allophycocyanin(APC): =A652-0.208(A615)/5.09 mg/ml
C- Phycoerythrin(PE): = A562-2.41(PC) - 0.849(APC)/9.62 mg/ml

RESULTS AND DISCUSSION

There has been a thirst of interest in the micro algal metabolites in recent years. This must be quenched off by continuous investigations for the value added products of blue green algae in cost effective way. In the present work, the protein and various pigments (Chlorophyll a, Carotenoids, Phycobilins) of the two blue green algae *Oscillatoria Sp.* and *Spirulina platensis* were studied.The cyanobacterial strains were isolated and confirmed microscopically. These strains were grown in synthetic medium invitro and the growth was attained in 7 to 9 days. The cells were then collected from medium and analyzed for protein, Chlorophyll a, Phycobilin and Carotenoid concentration.Blue green algae-cyanobacteria has been used for thousands of years as a good food source for human and animal due to the excellent nutritional profile and high carotenoid content[2,5,9].A new course has been developed in the feed business towards the use of natural ingredients and away from antibiotics, synthetic colours and other chemicals [26,3,19].Realizing the need to develop the production of natural product sources, recent years show a tremendous

interest in the field of algal biotechnology.In the present study, the physiochemical properties of the cyanobacterium *Oscillatoria Sp. and Spirulina platensis* were assayed by growing in synthetic medium. They were analyzed for protein chlorophyll carotenoid and Phycobilin concentration.

The protein concentration of *Oscillatoria Sp. and Spirulina platensis* grown invitro were 0.19mg/ml and 0.36 mg/ml.The Chlorophyll concentration of Spirulina was found to be 77.9mg/ml and that of *Oscillatoria Sp.*was 76.4mg/ml when grown invitro. Similarly the carotenoid concentration of *Spirulina platensis* was 0.024 mg/ml and that of *Oscillatoria Sp.* was 0.022 mg/ml .The Phycocyanin concentration of *Spirulina platensis* is 0.16 mg/ml and that of *Oscillatoria Sp.* show no Phycocyanin concentration the allophycocyanin concentration of *Spirulina platensis* was found to be at increased level of 6.3 mg/ml while that of *Oscillatoria Sp.* was 0.1mg/ml**.** According to Burja *et al*., (2002)[7], the variation or increase in the Surface area to volume ratio may influence the growth rate, wet weight and pigment production.Several studies were done on the Carotenoid, Chlorophyll and Phycobilin concentrations of *Spirulina platensis* [4,18,21] and *Oscillatoria Sp* [7,28].

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Table: 1.Comparison of protein and phytochemical concentration in *Spirulina platensis* and *Oscillatoria Sp.*grown *invitro*.

Phytochemical concentration	***Spirulina platensis* mg/ml**	***Oscillatoria Sp.*mg/ml**
Protein	0.36	0.19
Chlorophyll a	77.9	76.4
Carotenoid	0.02	0.02
Phycocyanin (PC)	0.16	0.0
Allophycocyanin (APC)	6.30	0.10
Phycoerythrin (PE)	0.0	0.08

RESEARCH ARTICLE

Green Synthesis of Silver Nanoparticles and Structural Elucidation of Root extract of *Hemidesmus indicus* R.Br. with Docking study for Anti Cancer activity.

Anis Fatima M.[1], B.Ramya[1]*, S.Vijikumar[2] and K.Ramanathan[3]

[1]Department of Biochemistry, JJ College of Arts and Science (Autonomous),Pudukkottai-622404, TamilNadu,India.

[2]TamilNadu Scientific Research Organisation, Pudukkottai-622003, TamilNadu, India.

[3]Department of Bioinformatics, Thanthai Hans Roever College of Arts and Science, Perambalur-621212, Tamil Nadu ,India.

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***Address for correspondence**
Mrs.B.Ramya
Assistant Professor, Departmet of Biochemistry,
JJ College of Arts and Science (Autonomous),
Pudukkottai-622404, TamilNadu, India.
E.mail: ramya_3406@yahoo.co.in

ABSTRACT

The medicinal plant *Hemidesmus indicus* R.Br. was selected for analysis and Characterization of its medicinal value based on phytochemical studies. The collected plants and their root parts were cleaned with tap water and dried under shade, then ground well to find powder. About 100gm of dry root powder of *Hemidesmus indicus* R.Br. was extracted with solvent ethanol using soxhlet apparatus at 60-70 degree temperature. Grayish black waxy residue was obtained. The residue was used for phytochemical analysis along with other qualitative assays such as UV-Vis, FT-IR, HPLC and XRD with synthesis of silvernano particles and Bioinformatic study for protein binding for anticancer activity. These results suggest that *Hemidesmus indicus* R.Br. might be a source of large amount of metabolites such as phenolics. Therefore, this result suggests that *Hemidesmus indicus* R.Br. extracts posses' compounds with anticancer activity.

Keywords: *Hemidesmus indicus* R.Br, soxhlet apparatus, UV-Vis, FT-IR, HPLC and XRD

INTRODUCTION

Hemidesmus indicus R.Br. (Family: Asclepiadaceae), commonly known as Indian sarsaparilla or Anantmool.It is a slender, laticiferous and twining shrub, occurs over the greater part of India. This plant is known in Ayurveda as a tonic, demulcent, diaphoretic, diuretic and blood purifier. It is beneficial in treating skin and urinary disorders. The plant is found throughout Central, Western and Southern part of India[2]. The coumarinolignoids hemidesmine and hemidesmin are the major chemical constituents of the herb, which give its therapeutic properties cools the urinary tract, which effectively alleviates the burning sensation during urinary infections and kidney disorders. As an antibacterial, treats skin diseases and promotes skin healing.*Hemidesmus indicus* R.Br. has antimicrobial, antioxidant, antihelminthetic and nervous stimulant activities, the present study aiming to envisage its effects by elucidating the structure of active metabolite [3,4,12].

MATERIALS AND METHODS

On the basis of its medicinal value which is available in the literature, *Hemidesmus indicus* R.Br. Plant roots were collected in Arimalam Village, Pudukkottai district, TamilNadu. They were taxonomically identified by local farmers, traditional healers and Prof.Palaniyappan, Department of Botany, JJ College of Arts and Science, Pudukkottai, TamilNadu, India and FRLHT ENVIS medicinal plant database website.

Preparation of plant extract for phytochemical studies

The root of *Hemidesmus indicus* R.Br. was dried under the shadow for 2 weeks, and it was made as coarsely powdered for extraction. About 100 gm of dry root powder of was *Hemidesmus indicus* R. Br. extracted with ethanol at 60^{o}c to 70^{o}c by continuous hot percolation using soxhlet apparatus. The extraction was filtered and kept in over at 50^{o}c for 24 hours to evaporate the extracts from them. A grayish black waxy residue was obtained. These extracts were used for phytochemical analysis qualitatively. Phytochemical analysis for major phytoconstituents of the plant extract was undertaken using standard qualitative methods as described by various authors along with UV-Visible spectroscopy, Infrared Spectroscopy, HPLC and X-ray diffraction methods.

Synthesis of Silvernano Particles

The 25 ml of 1M silver nitrate ($AgNO_3$) solution was taken in a conical flask and added 1.5 ml of *Hemidesmus indicus* R.Br. plant ethanolic root extract solution for reduction of silver nitrate into Ag^{+} ions and kept at room temperature. After around 12 hours the colour of the mixture turned into light purple and continuously changed to purple-brown indicating the formation of silver nanoparticles [1,5,13].

Study of Bioinformatics

The identified sequence for *Hemidesmus indicus* R.Br. was retrieved from Databank and submitted to GOR tool. The calculation of secondary structural parameters was done by GOR tool .The hydrophobic nature of the compounds were calculated and selected for docking with the receptor (Table.3,Figure1,2,3,4,5)

RESULTS AND DISCUSSION

Phytochemical constituents like alkaloids, flavoniods, carbohydrates, glycosides, phytosterols, fixed oil and fats, proteins, phenolic compounds, and saponins of ethanolic root extract *Hemidesmus indicus* R.Br. were analysed by qualitatively(Table1).

On the basis of UV-Vis and FT-IR spectral analysis on *Hemidesmus indicus* R.Br. we have found the following data. UV-Vis shown in graph 1 yielded 1 elevations (255.16 nm) and in graph 2 yielded 2 elevations 255.16 nm and 278.60 nm (silvernano particles of *Hemidesmus indicus* R.Br.) the values were interpreted with standared table values and confirm the presence of phenolics in the given sample. FT-IR is shown in graph 3 yielded Maximum peak level 3757.65$cm^{-1}$and Minimum peak level 434.04 cm^{-1}. FT-IR studies confirm the presence of functional groups in the compound listed in the table 2, So that the compound may be phenolics.The X-ray diffraction results clearly show that the silver nanoparticles formed by the reduction of Ag+ ions by the root extract of *H. indicus* are crystalline in nature. The sequence for *Hemidesmus indicus* R.Br. was retrieved from databank and submitted to GOR tool. The calculation of Secondary structural parameters was done by GOR tool. The hydrophobic nature of the compounds were calculated and selected for docking with the receptor.

Fimognari C.,2011 [2] stated that the molecular basis of the antileukemic effects of *Hemidesmus* and identify the mitochondrial pathways and [Ca(2+)] as crucial actors in its anticancer activity. On this basis, we conclude that *Hemidesmus* can represent a valuable tool in the anticancer pharmacology, and should be considered for further investigations. Ravishankara et al [8] studied antioxidant activity of methanolic extract of *H. indicus* root bark was evaluated in several in vitro and ex vivo models. Further, preliminary phytochemical analysis and TLC fingerprint profile of the extract was established to characterize the extract which showed antioxidant properties.Saravanan *et al* [9,10,11] studied that treatment *with H. indicus* extract offers protection against free radical-mediated oxidative stress in plasma, erythrocytes, and liver of animals with ethanol-induced liver injury, and his other data indicate that treatment with EHI(Ethonal extract of *H. indicus)* offers protection against free radical-mediated oxidative stress in kidney of animals with ethanol-induced nephrotoxicity.Prabakan*et al* [7]stated that oral treatment with the ethanol extract of *Hemidesmus indicus* roots (100 mg/kg, for 15 days) significantly prevented rifampicin and isoniazid-induced hepatotoxicity in rats Based on above discussion, in my research UV, FTIR, HPLC and XRD studies concluded that phenolics are antioxidant compound may be present in the root of *Hemidesmus indicus.* R.Br. Finally bioinformatics study was strongly evidenced for above mentioned activity particularly anticancer activity(Table 2, Graph1,2)[6].

CONCLUSION

In conclusion, these results suggest that *Hemidesmus indicus* R.Br. might be a source of large amount of metabolites such as phenolics. Therefore, this result may suggest that *Hemidesmus indicus* R.Br. extracts possess compounds with anticancer, antimicrobial and antioxidant properties which can be used as Phytochemical agents in new drugs for therapy of cancer disease and other ailments of human.

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Table 1: Depicts various phytochemicals analyzed qualitatively in ethanol extract of *Hemidesmus indicus* roots.

S.No.	Metabolites	Results
1	Alkaloids	Absent
2	Flavonoids	present
3	Terpenoids	Absent
4	Fixed oils	present
5	Phytosterols	present
6	Saponins	Absent
7	Phenolic compounds	present
8	Fats	present
9	Carbohydrates	present
10	Proteins	present
11	Glycosides	present
12	Tannins	present

Table 2: Composition of Structural Parameters

Parameters	Composition	Percentage
Alpha Helix	184	36.15
Extended Strand	100	19.65
Random Coil	225	44.20

Table 3: Calculation of Hydrophobic Nature for Compounds

Name of the Compound	Hydrophobicity
1-Methylnaphthalene	3.9
Fluorene	4.2
Butylatedhydroxyanisole	3.2
4-tert-Butylphenyl Salicylate	5.5
Butylatedhydroxytoluene	5.3
Phenanthrene	4.5

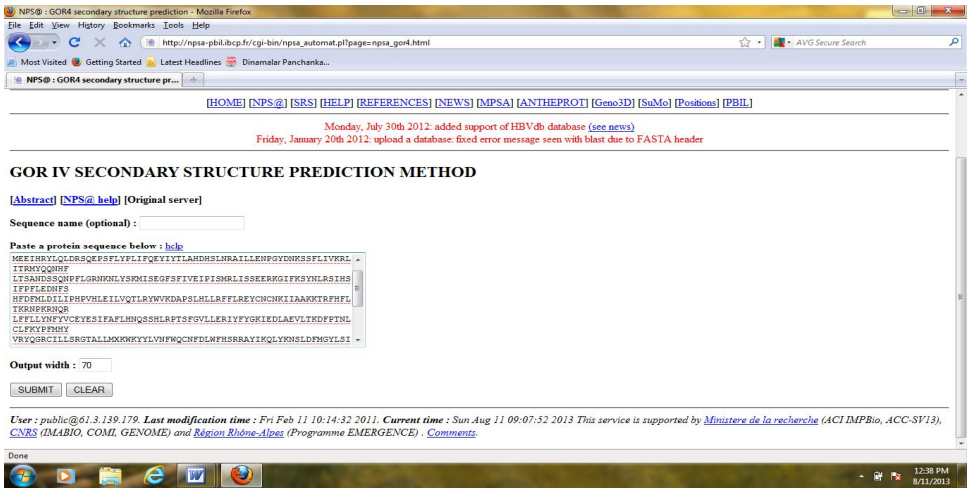


Figure 1: Submission of sequence in to GOR Tool

Figure 2: Calculation of Structural Parameters

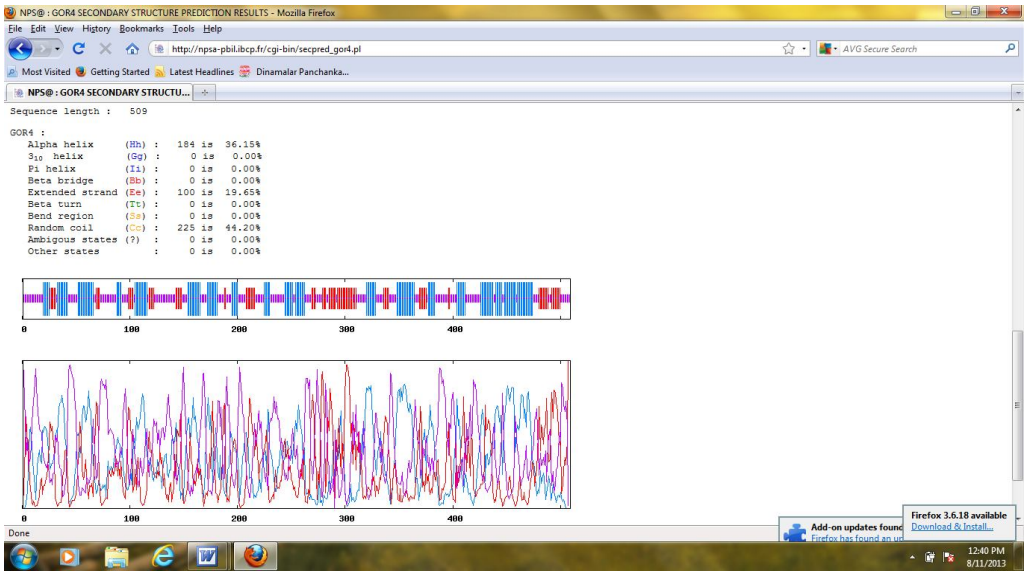


Figure 3: Composition of Structural Parameters

GOR4

Alpha helix (Hh) : 184 is 36.15%

3_{10} helix (Gg) : 0 is 0.00%

Pi helix (Ii) : 0 is 0.00%

Beta bridge (Bb) : 0 is 0.00%

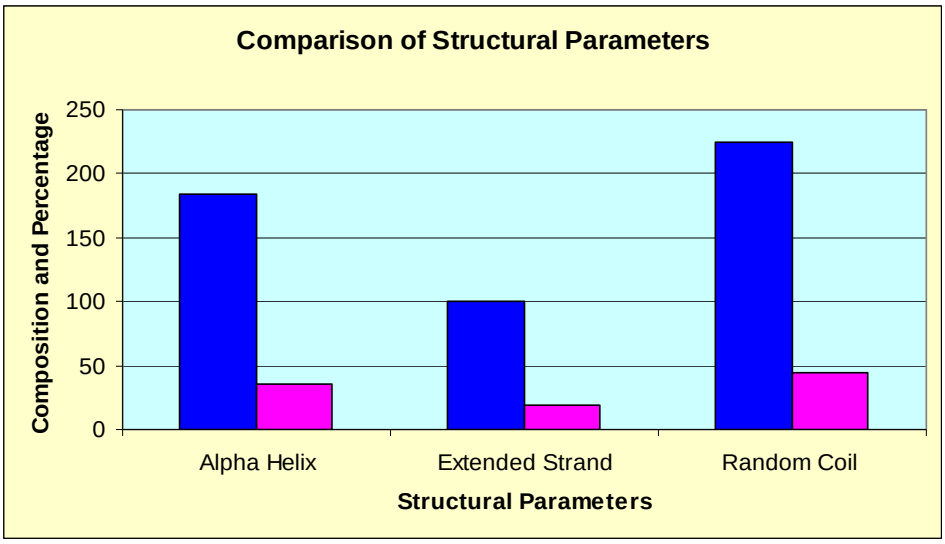
Extended strand (Ee) : 100 is 19.65%

Beta turn (Tt) : 0 is 0.00%

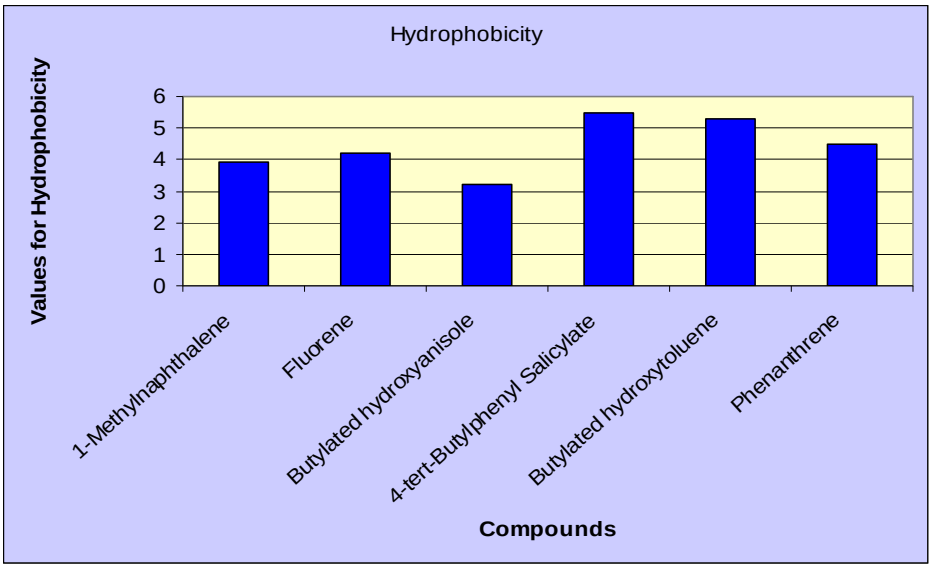
Bend region (Ss) : 0 is 0.00%

Random coil (Cc) : 225 is 44.20%

Other states : 0 is 0.00%



Graph 1: Comparison of Structural Parameters



Graph 2: Comparison of Compounds derived from *Hemidesmusindicus*R.Br.

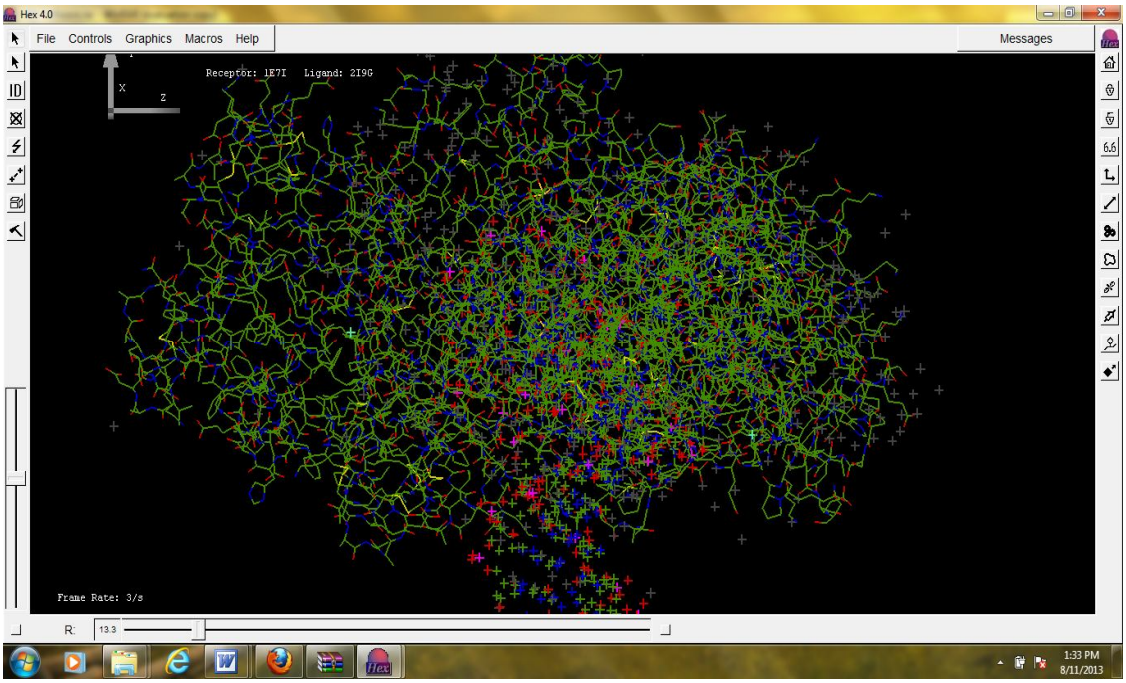


Figure 4: Before docking the receptor and ligand

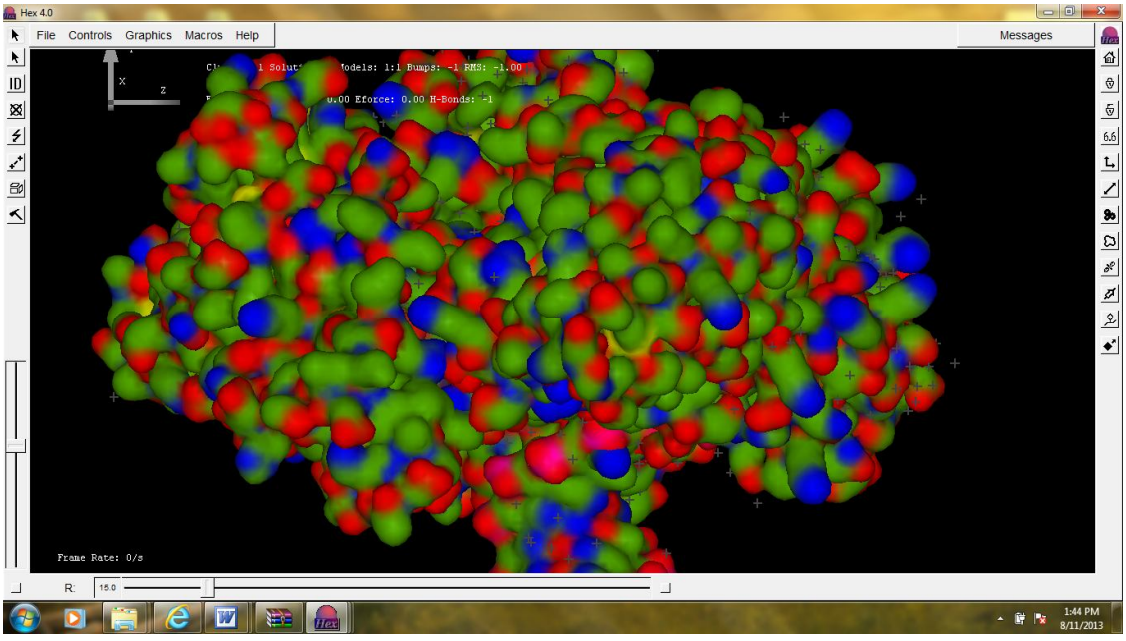


Figure 5: Docked structures of receptor and ligand

Docking Score - 75.13
Serum albumin - Phenanthrene

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